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8

9 **Abstract**

10 Host-pathogen co-evolution is central to shaping natural communities and is the
11 focus of much experimental and theoretical study. For tractability, the vast
12 majority of studies assume the host and pathogen interact in isolation, yet in
13 reality they will form one part of complex communities, with predation likely to
14 be a particularly key interaction. Here I present the first theoretical study to
15 assess the impact of predation on the coevolution of costly host resistance and
16 pathogen transmission. I show that fluctuating selection is most likely when
17 predators selectively prey upon infected hosts, but that saturating predation, due
18 to large handling times, dramatically restricts the potential for fluctuations. I also
19 show how host evolution may drive either enemy to extinction, and demonstrate
20 that while predation selects for low host resistance and high pathogen
21 infectivity, ecological feedbacks mean this results in lower infection rates when
22 predators are present. I emphasise the importance of accounting for varying
23 population sizes, and place the models in the context of recent experimental
24 studies.

25

26 **Keywords:** Host-pathogen, coevolution, communities, fluctuating selection

27 **Introduction**

28 Antagonistic co-evolution between hosts and their pathogens is central to
29 shaping the structure and function of biological communities [1,2]. A rich field of
30 experiment and theory has been developed to understand the drivers of host-
31 pathogen co-evolution and its impact on ecological dynamics [3-6]. However, for
32 tractability the vast majority of studies assume that the host and pathogen exist
33 in isolation. In reality host-pathogen interactions will be embedded within
34 complex communities with an array of biological interactions. These community
35 interactions will have significant impacts on the host-pathogen interaction,
36 which will in turn feed back to the community dynamics [6]. Predation will be
37 particularly significant due to the direct effects on host population size, as well as
38 indirect links between infection and predation. Classic empirical work has shown
39 that hosts with higher pathogen burdens are more likely to be predated [7,8],
40 potentially altering selection pressure on both antagonists, and thus impacting
41 the community structure itself.

42

43 Theoretical studies on the co-evolution of host resistance and pathogen
44 infectivity have found a range of possible qualitative outcomes, including long-
45 term stable investment (Continuously Stable Strategies), branching to
46 polymorphism and co-evolutionary cycles (fluctuating selection dynamic),
47 depending on the ecological and evolutionary context [9-20]. A particular focus
48 has been on fluctuating selection (FSD) given its importance to the maintenance
49 of diversity [21], evolution of sex [22] and local adaptation [23]. It is well known
50 that highly specific, 'matching-allele', infection mechanisms give rise to FSD due
51 to negative frequency-dependent selection [17,18], while gene-for-gene

mechanisms (variation between specialists and generalists) can lead to FSD if there are costs [19,20]. Recent work including explicit ecological dynamics found that cycles of host and pathogen investment could occur even without specificity [14]. However we have little understanding of how robust theoretical predictions are to including community interactions.

There is increasing awareness in experimental literature of the importance of community interactions to host-pathogen co-evolution [1,2,6], and there have been some direct experimental tests [24-26]. Friman & Buckling [24] found that the Arms Race Dynamic between a bacteria (*Pseudomonas fluorescens*) and its phage ($\Phi 2$) appeared to break down when a predatory protist (*Tetrahymena thermophila*) was present, while Örmälä-Odegrip et al. [26] found that selection due to predatory protists led to lower susceptibility to phage infection in both *Serratia marcescens* and *Pseudomonas fluorescens*. Alongside this experimental work, there is increasing theoretical focus on how the evolution of hosts and pathogens [27-32] are separately impacted by an immune predator (a predator that cannot be infected by the parasite). These studies have shown that pathogens invest in higher virulence and transmission when a predator is present [27], while hosts maximise defence to parasitism at intermediate predation rates [31]. In contrast to standard models, predation allows for evolutionary branching to coexistence in pathogens (if virulence and predation are linked; [28] v [33]) and the pathogen can be eradicated through host evolution ([30] v [34]). These studies provide a broad examination of the separate evolutionary properties of hosts and pathogens in the presence of a predator. However, given the importance of the co-evolutionary setting to the

potential for FSD [14,16-20], the differing predictions of the impact of predation on parasites [27] to hosts [31] and the importance of changing population sizes to host-parasite coevolution [5], it is vital that we investigate the full coevolutionary dynamics in the presence of a predator. Here I present a model of the co-evolution of host resistance (through reduced susceptibility) and pathogen transmission with non-specific infection, and respective costs to host birth rate and virulence.

Methods

I use a standard model of the population dynamics of susceptible (S) and infected hosts (I), adding an immune predator (P), as given by the following ordinary differential equations,

$$\begin{aligned} (1) \frac{dS}{dt} &= (b - qH)S - dS - \beta SI + \gamma I - c\rho(S, I)SP \\ (2) \frac{dI}{dt} &= \beta SI - (d + \alpha + \gamma)I - c\phi\rho(S, I)IP \\ (3) \frac{dP}{dt} &= ecP(S + \phi I)\rho(S, I) - \mu P \end{aligned}$$

Susceptible hosts reproduce at birth rate b which is reduced due to crowding by a factor q ($H=S+I$). All hosts die at natural death rate d . Transmission is a density-dependent term with coefficient β . As well as the natural death rate, infected hosts suffer an additional mortality, which I define as virulence, at rate α , and can recover back to susceptibility at rate γ . Both susceptible and infected hosts are at risk of predation with coefficient c , with a functional response given by $\rho(S, I) = 1/(1 + ch(S + \phi I))$ (see ESM and figure S1). If $h=0$ (i.e. there is no 'handling time'), the functional response is linearly dependent on the effective

host density, $S + \phi I$ (type I). If $h > 0$ then the response is saturating at higher effective host densities (type II). In what follows I assume the type I response unless otherwise stated. I also allow the predator to selectively predate infected hosts by the inclusion of the parameter $\phi > 1$. Predators convert energy from eating hosts in to births through parameter e , and die at rate μ . Note that I do not assume any link between virulence and predation, as in [28].

When there is linear (type I) predation, the full host-pathogen-predator equilibrium (where it exists) is always stable. However, for a type II response population cycles can occur. In the type I case, the resident equilibrium for \hat{S} and \hat{I} can be found as,

$$(4) \hat{S} = \frac{\alpha + d + \gamma}{\beta} + \frac{c\phi}{\beta} \hat{P}$$

$$(5) \hat{I} = \frac{\mu}{ec\phi} - \frac{\alpha + d + \gamma}{\beta\phi} - \frac{c}{\beta} \hat{P}$$

Therefore the susceptible density will always increase as the predator is introduced, while the infected density will always decrease (the total host density, \hat{H} , also decreases with increasing \hat{P}). Note that this relationship is independent of whether ϕ is greater than or less than unity. This is because, as in classic host-parasite models, the susceptible density is regulated by the parasite [35]. Therefore the increase in predation ultimately benefits susceptible hosts by reducing the density of infecteds. Models with different underlying assumptions, such as an explicit carrying capacity in the host [36], may yield different feedbacks.

I assume that the host can evolve its susceptibility to infection, and the pathogen its infectivity. As such I need to determine how the two jointly control transmission. Here I use a multiplicative function, $\beta(\sigma, \tau) = \sigma\tau + k$, where σ is the host's susceptibility and τ the pathogen's transmission. Such a 'universal' infection function has been commonly used in theoretical studies [11,12,15,17], and is representative of systems where infection is not specific to certain combinations of host and parasite strains [37-39]. I assume that investment in lower susceptibility and higher transmission incur respective costs for the host (lowered birth rate) and pathogen (increased virulence). Examples of the trade-offs are plotted in figure S2; see ESM and figure legends for the form of the trade-off functions. I model co-evolution using the evolutionary invasion analysis framework of adaptive dynamics [40-42], assuming that small, rare mutants (σ_m, τ_m) arise and attempt to invade a resident equilibrium. The success of the mutant is given by its invasion fitness, which is defined as its growth rate whilst rare. As described in the online ESM, assuming a type I functional response, this is given for the host by,

$$(7) s(\sigma_m; \sigma, \tau) = (T + \sqrt{T^2 - 4D})/2$$

where,

$$\begin{aligned} T &= b(\sigma_m) - q\hat{H} - 2b - \beta(\sigma_m, \tau)\hat{I} - c(1 + \phi)\hat{P} - \alpha(\tau) - \gamma \\ D &= -(b(\sigma_m) - q\hat{H} - b - \beta(\sigma_m, \tau)\hat{I} - c\hat{P})(b + \alpha(\tau) + \gamma\beta + c\phi\hat{P}) - \gamma\beta(\sigma_m, \tau)\hat{I} \end{aligned}$$

and for the pathogen,

$$(8) r(\tau_m; \sigma, \tau) = \beta(\sigma, \tau_m)\hat{S} - (d + \alpha(\tau_m) + \gamma) - c\phi\hat{P}$$

where all population densities are evaluated at the resident equilibrium (denoted by hats).

Assuming small mutations, the co-evolutionary dynamics of the traits σ and τ over evolutionary time can then be approximated by a pair of ordinary differential equations [42] (see ESM),

$$(9) \frac{d\sigma}{dT} \propto \hat{S} \frac{\partial s}{\partial \sigma_m} \Big|_{\sigma_m=\sigma}$$

$$(10) \frac{d\tau}{dT} \propto \hat{I} \frac{\partial r}{\partial \tau_m} \Big|_{\tau_m=\tau}$$

The possible long-term outcomes are: (1) a Continuously Stable Strategy (CSS) in both antagonists where the host and pathogen both invest in a stable level of investment, (2) co-evolutionary cycles (FSD), (3) evolutionary branching in one or both species, (4) maximisation/minimisation to the imposed (physiological) limits of the trait by one or both species. In the latter two cases, one species may exhibit this outcome, while the other could exhibit any of behaviours 1, 3 or 4 [14]. Further details of the methods are given in the online ESM.

Results

Qualitative outcomes

In figure 1 I show the qualitative outcome from simulations as the host and pathogen trade-off curvatures (p_h and p_p) are varied, for (a) linear (type I), and (b)-(d) saturating (type II) predation ($h=0.4, 0.45, 0.5$). Note that accelerating (increasingly costly) trade-offs occur for $p_h > 0$ but $p_p < 0$ (marked '*acc.*' in figure 1; see also figure S2). A range of qualitatively different outcomes are possible (see sample outputs in figure S3). In all cases, while the pathogen's trade-off is accelerating, if the host's trade-off is also accelerating there is a coevolutionary CSS, while if the host's trade-off decelerates the host branches (and the parasite remains at its CSS). The potential for cycles (FSD) and pathogen branching

depend on the handling time. For type I predation (fig 1a), if both trade-offs decelerate (marked '(dec.)'; $p_h < 0, p_p > 0$) then FSD is common. Initially introducing a handling time (fig 1a vs 1b) shifts the region of FSD to higher parasite trade-off curvatures but any host trade-off shape, suggesting the parasite trade-off must be reasonably decelerating for selection to be destabilised. This also introduces greater regions of pathogen branching, either on its own or together with the host. However, figures 1(b)-(d) show that cycles rapidly disappear once the handling time reaches a threshold value (here between $h=0.4$ and $h=0.5$). Comparing these figures the cycles are lost in two ways. First, the dynamics can be stabilized towards an evolutionary branching point, generally resulting in both species branching. Alternatively, the predator can go extinct during the cycle (after this the host maximizes susceptibility and the pathogen minimizes infectivity). The irregular nature of these transitions (their 'scattered' nature) is due to small stochastic variations between simulations –small amplitude cycles being close enough to a singular point to branch, or low predator densities during a cycle being approximated to zero.

Why does saturating predation cause coevolution to stabilise towards a branching point? When predation is linear, mortality is higher (figure S1). With selective predation of infecteds, this will strengthen selection for host resistance, pushing host investment, temporarily, to higher levels and continuing the cycles. When predation saturates and mortality is lower, this effect is reduced and the dynamics are stabilized.

Figure 2 shows how FSD depends on the predation rate, c , and selective predation, ϕ . Here we see that FSD is most common when there is high selective

predation but low general predation. This means that infected hosts suffer much higher mortality than susceptible hosts, fitting with the above argument that this increases selection for host resistance, thus destabilizing selection. This region is bounded on both sides by regions where one or both species branches. We also see that when both selective and general predation are low, the predator dies out and when both are high the pathogen dies out.

Extinction of the predator or pathogen

Invasion/exclusion thresholds exist for the pathogen and predator ([30]; see ESM). This allows for one of the species to be driven to extinction. A particularly interesting example of pathogen extinction can be seen in the phase portrait of figure 3, highlighting regions where the pathogen (red) or predator (blue) cannot persist (a case of predator extinction is in figure S4). The solid line shows a trajectory that tends to intermediate host and high pathogen investment when all three species coexist (blue dot). However, changing only the initial condition, the dashed line crosses the threshold for pathogen persistence, at which point the pathogen goes extinct. Note that this extinction occurs due to the host increasing its susceptibility to infection, a rather unintuitive result. This occurs because increasing susceptibility leads to a greater predator density, pushing the infected host population to ever lower densities. Again, note that increased predator density always leads to increased susceptible and decreased infected densities, regardless of selective predation.

Continuously Stable Strategies

Figure 4 explores how predation impacts host and pathogen investment at a Continuously Stable Strategy (CSS). Figure 4a shows the host (solid) and pathogen (dashed) strategies as predation rate, c , is varied, with the overall transmission coefficient, β , in figure 4b and the resulting *per-capita* rates of infection, $\beta\hat{I}$, and predation, $c\hat{P}$, in figure 4c. For low predation the predator cannot persist and there is a fixed level of investment. Once the predator can persist, the pathogen increases its investment, while the host displays a 'U'-shaped curve (fig 4a), leading to an overall increase in the transmission coefficient (fig 4b). However, fig 4c shows that the negative feedback from predation to the infected density means that the *per-capita* rate of infection, $\beta\hat{I}$, is significantly reduced. Thus high rates of predation lead to high host susceptibility and high pathogen infectivity, yet relatively low rates of infection in the population. Similar patterns are found for varying other parameters (figure S5).

Evolutionary branching

Purely host-parasite models with ecological dynamics and universal transmission have found that branching can occur such that two hosts and one pathogen, or two of each antagonist, coexist [12,15]. Further work found that adding a predator means the pathogen can branch against a monomorphic host when there is a link between virulence and predation [28]. Here, I find the stronger result that the pathogen can branch (against a monomorphic host) even without this link when predation saturates (figures 1,2). This indicates the emergence of a negative feedback to pathogen selection once predation is saturating. Further branching is not possible and the maximum level of diversity

remains two hosts-two pathogens. After the pathogen has branched, the system stabilizes. In particular, the predator cannot be driven to extinction without one of the pathogen strains first being excluded (since standard host-parasite models cannot support two pathogen strains [12]). Examining simulation results, after host branching it seems there is never extinction of either the predator or pathogen.

Discussion

There is increasing focus on understanding how community interactions impact host-pathogen co-evolution [1,2,6]. I have examined the co-evolution of host resistance (reduced susceptibility) and pathogen transmission, with respective costs to birth rate and virulence, in the presence of a predator. Fluctuating selection (FSD) is a particularly important co-evolutionary behaviour since it is the only sustained dynamic outcome in a constant environment, and is the focus of much theoretical study [14,16-20]. I have found that while FSD is common when the predator's functional response is linear, if predation saturates at high host densities FSD becomes an increasingly rare outcome, with evolutionary branching of the pathogen occurring instead. FSD is also promoted when there is strong selective predation of infected hosts. The driver of both results is that mortality of infected hosts is higher when predation is selective and does not saturate, destabilizing selection near an evolutionary attractor. Thus host-pathogen FSD may be expected in communities with highly selective predators with low handling times. In an experimental study of a microbial system the addition of a predatory protist appeared to breakdown an Arms Race Dynamic, but there was no conclusive evidence that the dynamics shifted to FSD [24]. It

would be interesting to conduct explicit experimental tests of how host-pathogen systems that exhibit FSD behave when a predator is added.

In standard models hosts cannot cause pathogen extinction through the evolution of costly resistance [34], but can when a predator is present [30]. Here I have shown a particularly unintuitive example of pathogen extinction caused by the host lowering its resistance. This drives an ecological feedback whereby the predator density increases and pushes pathogen numbers to extinction. It is notable that there is no evolutionary rescue of the pathogen. This is in fact intuitive since as the pathogen numbers decrease the relative speed of mutation also decreases. Host-driven pathogen extinction, in the absence of predation, has been found in experimental studies when further pressures, for example, population bottlenecks [43] or reduced resource availability [44], are placed on the pathogen. This appears consistent with the result that extinction may occur when a predator is introduced. Intriguingly, in their experimental study of bacteria-phage co-evolution in the presence of a predatory protist, Friman & Buckling [24] report a case of phage being driven to extinction, and it would be fascinating to see whether such a result is repeated elsewhere.

I have shown that while the introduction of a predator may lead to lower host resistance and higher pathogen infectivity at a co-evolutionary CSS compared to when no predator is present, the negative feedback from predators to the infected density means that there are in fact lower *per-capita* rates of infection than when the predator is absent. This has important consequences for how infection rates are measured in empirical studies, suggesting opposing patterns

of infection may be predicted depending on whether population sizes are controlled or not. Previous theory has shown, when only one antagonist evolves, that the pathogen should increase transmission when a predator is added [27] but the host should maximise defence at intermediate predation rates [31]. These results remain broadly true here, but give a misleading impression of the full co-evolutionary outcome when feedbacks to population sizes are not included. Interestingly, experimental results from two bacteria-phage-protist systems found hosts exhibited lower susceptibility to phage infection when a predatory protist was present [26]. This host response is consistent with the results here and earlier [31] assuming predation rates are not too high or co-evolution and ecological feedbacks are not fully present. More generally, the prediction here that overall infection rates may be lower when a predator is present is consistent with two key experimental studies [24,26]. Interestingly, Friman & Buckling [24] also reported that the introduction of the protist lowered overall host numbers, as would be expected here. It would be interesting to see whether direct experimental tests in the presence and absence of predators, including measures of population sizes, confirm the findings here.

Almost all natural and managed populations are part of communities, and this work is likely to have important implications to understanding a range of empirical systems, not least in microbial communities [2,4,45,46]. However, understanding antagonistic co-evolution in the context of complex communities is still an emerging field, and many open questions remain. For example, here I assumed no specificity in infection. Previous theory has shown that such specificity has implications for both static and transient diversity [14,15], and

this may be more realistic for modelling certain systems. Further, I have assumed that the additional interaction is with an immune predator, but other interactions, such as mutualisms or competitors, may lead to different feedbacks. A broader assessment of the impacts of community interactions on antagonistic coevolution should be a long-term goal of both experiment and theory [6].

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Competing Interests

I have no competing interests.

Ethics

No ethics approval was required.

Data Accessibility

C++ code for the simulations is available as ESM.

Author Contributions

AB is the sole contributor.

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Figure Legends

Figure 1: Qualitative output from numerical simulations of the co-evolutionary dynamics for differing handling times, (a) $h=0$, (b) $h=0.4$, (c) $h=0.45$, (d) $h=0.5$, as the shape of the host and parasite tradeoffs vary. Accelerating ('acc.') and decelerating ('dec.') trade-offs are highlighted on the plots. The simulations were run (see ESM) and the output analysed and classified. CSS=Continuously Stable Strategy, BR=Branching, MX=Maximisation of trait, MN=Minimisation of trait, FSD=Fluctuating Selection/Cycles. See colorbar for classifications.. Parameter values: $q = 0.5, d = 0.2, \gamma = 0.2, \phi = 3, k = 0.5, \mu = 0.5, c = 0.15$. The trade-offs, linking transmissibility and virulence in the pathogen, and susceptibility and birth rate in the host, are given by $\alpha(\tau) = 1.06 - \frac{1-\tau}{1+p_p\tau}$, $b(\sigma) = 1.92 + \frac{0.16\sigma}{1+p_h(\sigma-1)}$ where p_p and p_h are varied along the x- and y-axes respectively.

Figure 2: Qualitative output from numerical simulations as the predation rate, c , and selective predation, ϕ , are varied. Parameters are as of figure 1 with $p_h = -0.5, p_p = 0.5$. See colorbar in figure 1 for classifications.

Figure 3: Phase portrait of co-evolution showing regions where the pathogen (red) or predator (blue) cannot persist. Parameter values are as of figure 1a, except $\phi = 2.25, k = 0.35$. The trade-offs are $\alpha(\tau) = 1.56 - \frac{1(1-\tau)}{1-0.23\tau}$, $b(\sigma) = 1.87 + \frac{0.21\sigma}{0.59+0.41\sigma}$.

481 **Figure 4:** How the co-CSS varies with predation, c . (a) Host, σ (solid) and
482 pathogen, τ (dashed) strategies, (b) transmission coefficient, β , and (c) per-
483 capita rate of infection, $\beta\hat{I}$ (solid) and predation, $c\hat{P}$ (dashed). Parameter values
484 are as in figure 1a with $p_p = -0.25$, $p_h = 0.25$.
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